

New Triterpenoids of *Mallotus repandus*

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Three new triterpenoids—3 α -hydroxy-13 α -ursan-28,12 β -olide 3-benzoate (**1**), 3 α -hydroxy-28 β -methoxy-13 α -ursan-28,12 β -epoxide 3-benzoate (**2**), and 3 α -hydroxy-13 α -ursan-28-oic acid (**3**)—and four known compounds were isolated from the stem and root bark of *Mallotus repandus*.

In Taiwan, the stems of *Mallotus repandus* (Willd.) Mueller-Arg. (Euphorbiaceae) have been used as an anti-inflammatory drug. Diterpenoids, triterpenoids, bergenin, cyano- γ -pyridone, and tannins have been reported to occur in this plant.^{1,2} In a search for bioactive constituents from this plant, three new triterpenoids, 3 α -hydroxy-13 α -ursan-28,12 β -olide 3-benzoate (**1**); 3 α -hydroxy-28 β -methoxy-13 α -ursan-28,12 β -epoxide 3-benzoate (**2**); and 3 α -hydroxy-13 α -ursan-28-oic acid (**3**), and four known compounds, 3-oxo-13 α -ursan-28,12 β -olide; 3 α -hydroxy-13 α -ursan-28,12 β -olide; ursolic acid; and bergenin, were isolated from the stem and root bark of *M. repandus*. In the present paper the characterization of **1**, **2**, and **3** is reported.

The high-resolution MS of **1** revealed [M]⁺ at *m/z* 560.3865, which corresponded to the molecular formula C₃₇H₅₂O₄. Its IR spectrum showed absorption bands characteristic of an ester group, a δ -lactone function, and an aromatic ring (1600 cm⁻¹). The ¹H and ¹³C NMR spectral features of **1** suggested an ursane triterpene. The ¹H NMR spectrum of **1** indicated signals for two secondary and five tertiary methyl groups, an axial methine proton at δ 4.80 on C-3 bearing a OCOR group, an equatorial methine proton at δ 3.93 on C-12 adjacent to the oxygen function of the lactone ring, and a phenyl group.² In addition, the presence of two methine carbon signals at δ 75.6 and 83.4, and two carbonyl carbon signals at δ 166.4 and 176.9, suggested that **1** was a lactone of an ursane triterpene similar to 3 β -hydroxy-13 α -ursan-28,12 β -olide 3-benzoate except that it exhibited different chemical shifts of H-3 and C-3 from those of 3 β -hydroxy-13 α -ursan-28,12 β -olide 3-benzoate.² Thus, **1** was characterized as 3 α -hydroxy-13 α -ursan-28,12 β -olide 3-benzoate. The ¹³C NMR assignments of **1** (Table 1) were made by performing ¹H-decoupled, DEPT, 2D NMR correlation experiments and by comparing the corresponding data for ursan-3 α ,19 α ,20 β -triol and methyl ursolate.^{3,4}

The HR FABMS of **2** revealed [M - 1]⁺ at *m/z* 575.4088, which corresponded to the molecular formula C₃₈H₅₅O₄. Its IR spectrum showed absorption bands characteristic of an ester group and an aromatic ring. The ¹H and ¹³C NMR data of **2** also suggested an ursane triterpene. The ¹H NMR spectrum of **2** was similar to that of **1**, except for absence of a signal at δ 3.93 (corresponding to H-12 α in **1**) and contained signals for a methine proton at δ 3.16, an aliphatic proton at 5.25 (s), and methoxyl protons at δ 3.37. The ¹³C NMR spectrum of **2** (Table 1) is also similar to that of **1**, except for the absence of signals at δ 83.4 and 176.9

Table 1. ¹³C NMR Chemical Shift Values of **1**, **2**, and **3**^{a,b}

carbon	1 (CDCl ₃)	2 (CDCl ₃)	3 (CDCl ₃)
1	40.5	41.1	37.6
2	21.6	21.3	29.0
3	75.6	75.5	72.6
4	38.0	37.2	38.4
5	57.4	59.7	54.3
6	19.3	19.4	19.2
7	32.4	32.6	34.2
8	38.5	38.7	39.3
9	49.8	52.7	49.0
10	37.2	37.1	38.4
11	39.4	37.5	20.7
12	83.4	76.6	29.0
13	39.1	41.1	44.3
14	35.9	34.2	40.3
15	27.9	28.3	29.3
16	32.4	32.2	33.7
17	51.4	40.7	49.0
18	49.8	50.2	54.2
19	39.0	41.3	39.1 ^c
20	39.0	41.3	39.2 ^c
21	30.0	29.9	37.2 ^d
22	31.5	35.7	36.9 ^d
23	14.3	14.6	15.0
24	34.6	35.9	31.1
25	23.3	21.0	23.3
26	20.4	24.0	31.6 ^d
27	30.5	30.6	35.8
28	176.9	101.5	180.4
29	18.0	17.6	19.2
30	10.1	10.1	10.4
1'	166.4	166.3	
2'	130.9	130.9	
3'	129.5	129.5	
4'	128.3	128.3	
5'	132.7	132.7	
6'	128.3	128.3	
7'	129.5	129.5	
OMe		56.0	

^a The number of protons directly attached to each carbon was verified by DEPT experiment. ^b Signals obtained by ¹H-¹H COSY, HMQC, HMBC, and NOESY techniques. ^c Assignments may be interchangeable. ^d Assignments may be revised.

(corresponding to C-12 and C-28 in **1**) and presence of signals for a methoxyl carbon at δ 56.0, an oxygenated carbon at δ 76.6, and an oxygenated tertiary carbon at δ 101.5. This suggested that the C, D, and E rings of **2** are different from those of **1**. In the HMBC spectrum, the carbon signal at δ 101.5 was correlated with the methine proton signal at δ 3.16 and the methoxyl proton signal at δ 3.37, respectively, and the aliphatic proton signal at δ 5.25 was correlated with the methoxyl carbon at δ 56.0 (Figure 1). The NOESY spectrum of **2** indicated correlations between H-12, H-13, and H-28 and Me-27, H-13, and H-28

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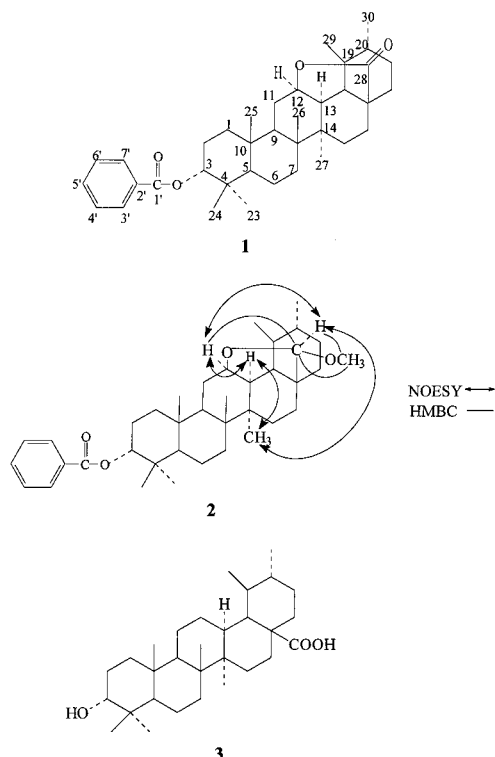


Figure 1. Structures of **1**, **2**, and **3**, and partial C/H long-range correlations and NOESY interactions of **2**.

(Figure 1). Thus, **2** was characterized as 3 α -hydroxy-28 β -methoxy-13 α -ursan-28,12 β -epoxide 3-benzoate. The ^{13}C NMR assignments of **2** (Table 1) were made by performing ^1H -decoupled, DEPT, 2D NMR correlation experiments and by comparing the corresponding data for **1**.

The HRMS of **3** revealed $[\text{M}]^+$ at m/z 458.3766, which corresponded to the molecular formula $\text{C}_{30}\text{H}_{50}\text{O}_3$. Its IR spectrum showed absorption bands characteristic of hydroxyl and carboxylic groups. The ^1H and ^{13}C NMR signals of **3** also indicated an ursane skeleton. The ^1H NMR spectrum of **3** contained signals for two secondary and five tertiary methyl groups, and an axial methine proton at δ 3.26 on C-3 bearing a hydroxyl group. The absence of any signal beyond δ 3.26 and any band near 1600 cm^{-1} in the IR spectrum suggested the saturated nature of this compound. Based on the above evidence, the chemical shift of H-3 and C-3 in the ^1H and ^{13}C NMR spectra of **3**, respectively, are identical to those of corresponding data for 3 α -hydroxy-13 α -ursan-28,12 β -olide, and the NOESY spectrum of **3** indicated correlation between H-13 (δ 1.89, m) and Me-27 (δ 0.95, s). Thus, compound **3** was characterized as 3 α -hydroxy-13 α -ursan-28-oic acid (Figure 1). The ^{13}C NMR assignments of **3** (Table 1) were made by performing ^1H -decoupled, DEPT, 2D NMR correlation experiments and by comparing the corresponding data for ursan-3 α ,19 α ,20 β -triol and methyl ursolate.^{3,4}

Experimental Section

General Experimental Procedures. Melting points are reported uncorrected. Optical rotation was obtained on a JASCO model DIP-370 digital polarimeter; UV spectra were obtained on a JASCO model 7800 UV-vis spectrophotometer; IR spectra were recorded on a Hitachi model 260-30 spectrophotometer; ^1H (400 MHz) and ^{13}C NMR (100 MHz) spectra

were recorded on a Varian Unity-400 spectrometer; and MS were obtained on a JMS-HX mass spectrometer.

Plant Material. The stem and root bark of *Mallotus repandus* (1.4 kg) were collected at Kaohsiung Hsien, Taiwan, during July 1996, and a voucher specimen has been deposited in the authors' laboratory (voucher no. Ma-01).

Extraction and Isolation. The stem and root bark (1.4 kg) of *M. repandus* were chipped and extracted with Me_2CO at room temperature. The Me_2CO extract was chromatographed over Si gel. Elution with cyclohexane-EtOAc (10:1) yielded **1** (50 mg); cyclohexane- CHCl_3 (9.5: 0.5) yielded 3-oxo-13 α -ursan-28,12 β -olide; cyclohexane- CHCl_3 (4: 1) yielded **2** (8 mg); CHCl_3 yielded **3** (10 mg) and 3 α -hydroxy-13 α -ursan-28,12 β -olide (15 mg); CHCl_3 -MeOH (9: 1) yielded ursolic acid (5 mg); and EtOAc-MeOH (9: 1) yielded bergenin (110 mg). These known compounds were identified by spectroscopic methods and comparison with authentic data² or sample.

3 α -Hydroxy-13 α -ursan-28,12 β -olide 3-benzoate (1**):** colorless needles (MeOH); mp 260–262 $^\circ\text{C}$; $[\alpha]_D^{27} -33^\circ$ (CHCl_3 , c 1.0); IR ν_{max} (KBr) cm^{-1} 1734, 1715; ^1H NMR (CDCl_3 , 400 MHz) δ 0.81 (3H, d, $J = 6.4$ Hz, Me-30), 0.87 (3H, d, $J = 6.4$ Hz, Me-29), 0.88 (3H, s, Me-23), 0.98 (3H, s, Me-27), 0.93 (3H, s, Me-24), 1.19 (3H, s, Me-26), 1.20 (3H, s, Me-25), 1.79 (1H, m, H-13 α), 3.93 (1H, m, $W_{1/2} = 9$ Hz, H-12 α), 4.80 (1H, m, $W_{1/2} = 16$ Hz, H-3 β), 7.36 (2H, m, H-4' and -6'), 7.47 (1H, m, H-5'), 7.96 (2H, m, H-3' and -7'); ^{13}C NMR (CDCl_3 , 100 MHz), see Table 1; EIMS (70 ev) m/z 560 $[\text{M}]^+$ (7), 455 $[\text{M} - \text{C}_6\text{H}_5\text{CO}]^+$ (4), 438 $[\text{M} - \text{C}_6\text{H}_5\text{COOH}]^+$ (15), 423 $[\text{M} - \text{Me}]^+$ (7), 370 (9), 314 (15), 187 (10), 123 (17), 105 (100); HREIMS m/z 560.3865 (calcd for $\text{C}_{37}\text{H}_{52}\text{O}_4$, 560.3855).

3 α -Hydroxy-28 β -methoxy-13 α -ursan-28,12 β -epoxide 3-benzoate (2**):** colorless powder (MeOH- CHCl_3); mp 155–157 $^\circ\text{C}$; $[\alpha]_D^{27} 179^\circ$ (CHCl_3 , c 0.05); IR ν_{max} (KBr) cm^{-1} 1716, 1603; ^1H NMR (CDCl_3 , 400 MHz) δ 0.77 (3H, d, $J = 6.4$ Hz, Me-30), 0.79 (3H, d, $J = 6.4$ Hz, Me-29), 0.84 (3H, s, Me-23), 0.86 (3H, s, Me-24), 0.90 (3H, s, Me-27), 1.00 (3H, s, Me-26), 1.03 (3H, s, Me-25), 1.75 (1H, m, H-13 α), 3.16 (1H, m, $W_{1/2} = 9$ Hz, H-12 α), 3.37 (3H, s, OMe), 4.81 (1H, m, $W_{1/2} = 16$ Hz, H-3 β), 5.25 (1H, s, H-28 α), 7.36 (2H, m, H-4' and -6'), 7.47 (1H, m, H-5'), 7.96 (2H, m, H-3' and -7'); ^{13}C NMR (CDCl_3 , 100 MHz), see Table 1; FABMS (negative) m/z 575 $[\text{M} - 1]^-$ (0.2), 545 $[\text{M} - \text{OMe}]^-$ (0.3), 455 $[\text{M} - \text{Me} - \text{C}_6\text{H}_5\text{CO}]^-$ (0.2), 423 $[\text{M} - \text{Me} - \text{C}_6\text{H}_5\text{COOH}]^-$ (1), 395 $[\text{M} - \text{CO}]^-$ (1), 189 (4), 123 (11), 105 (34), 69 (90), 55 (100); HRFABMS m/z 575.4088 (calcd for $\text{C}_{38}\text{H}_{54}\text{O}_4$, 575.4100).

3 α -Hydroxy-13 α -ursan-28-oic acid (3**):** colorless needles (CHCl_3); mp 155–157 $^\circ\text{C}$; $[\alpha]_D^{27} +53^\circ$ (CHCl_3 , c 0.05); IR ν_{max} (KBr) cm^{-1} 3339, 1688; ^1H NMR (CDCl_3 , 400 MHz) δ 0.79 (3H, s, Me-23), 0.87 (3H, d, $J = 6.4$ Hz, Me-30), 0.89 (3H, d, $J = 6.4$ Hz, Me-29), 0.95 (3H, s, Me-27), 1.01 (3H, s, Me-24), 1.14 (3H, s, Me-25), 1.23 (3H, s, Me-26), 1.83 (m, H-13 α), 3.26 (1H, m, $W_{1/2} = 16$ Hz); ^{13}C NMR (CDCl_3 , 100 MHz), see Table 1; EIMS (70 ev) m/z 458 $[\text{M}]^+$ (0.3), 440 $[\text{M} - \text{H}_2\text{O}]^+$ (2), 395 $[\text{M} - \text{COOH}]^+$ (1), 307 (21), 203 (11), 189 (14), 175 (23), 163 (22), 152 (39), 123 (48), 107 (54), 95 (80), 55 (100); HREIMS m/z 458.3766 (calcd for $\text{C}_{30}\text{H}_{50}\text{O}_3$, 458.3759).

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